

hGH ADA ELISA Catalog EL-141-878

For the determination antibodies to hGH in serum and plasma.

Introduction

The AffinityImmuno hGH ADA ELISA kit contains the components required for measuring antibodies to hGH in biological matrices such as human serum, plasma or cell culture supernatant.

Principle of the assay

This assay employs the indirect enzyme immunoassay technique. hGH is coated onto a 96 well microplate. Calibrator and test samples prepared by dilution into assay buffer and are pipetted into the appropriate wells. Antibodies to hGH present in biological matrices are bound to the immobilized hGH. After washing away any unbound substances, antibodies antibody-enzyme reagent and a substrate solution is added to the wells for color development. The color development is proportional to the amount of antibodies to hGH present in test samples. The color development is stopped and the intensity of the color is measured.

Materials and storage

Store kit components at -20°C unless specified otherwise. DO NOT USE past kit expiration date. Some vials contain a small amount of reagents. Spin tubes on pulse setting prior to opening.

Each kit includes:	Count
Coated microtiter plate, 96 wells (1x8 strips)	1
Calibrator diluent	1.8 ml
Calibrator (1mg/mL)	12 µl
10X wash buffer	50 ml
Assay buffer	50ml
1000X detection reagent	17µl
TMB	12 ml
TMB stop solution	12 ml
Do not mix or substitute reagents with those from other lots.	

Materials and instruments required but not supplied

- Precision pipettes calibrated to deliver 5-1000µL
- Multi-channel pipette calibrated to deliver 50-200µL
- Plate shaker
- Disposable tips
- Vortex-Mixer
- Distilled or de-ionized water
- Microplate reader capable of reading 450nm with background subtraction at 620nm

Safety precautions

- The test protocol must be followed strictly.
- All reagents containing human material should be handled as if potentially infectious. Operators should wear gloves and protective clothing when handling any patient sera or serum based products.
- The kit reagents contain antimicrobial agents, acid and 3,3',5,5'-tetramethylbenzidine. Avoid contact with the skin and eyes. Rinse immediately with plenty of water if any contact occurs.
- Any liquid that has been brought into contact with potentially infectious material has to be discarded in a container with a disinfectant. Disposal must be performed in accordance with local regulations.
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- Only trained laboratory personnel should execute this test.

Preparation of reagents

Prepare only the appropriate amount of required reagent on the day of use. Store all reagents as per instructions stated on the label.

- 1. Wash Buffer (1X) Preparation:** Dilute wash buffer concentrate with ultra-pure water 1/10 before use (for example add 50mL concentrate to 450mL ultra-pure water). Mix well.
- 2. Detection Reagent (1X) Preparation:** Dilute detection reagent with assay buffer 1/1000 before use (for example add 11µl concentrate to 11ml of assay buffer). Mix well.
- 3. Calibrator Preparation:** Dilute the calibrator from 1mg/ml down to 10µg/ml by pipetting 5µl of calibrator stock into 495µl assay buffer. Label "Cal. Int." Mix well. Prepare calibrators with concentrations ranging from 1920 ng/ml to 30 ng/ml. The following is an example calibrator curve.

Sol'n ID	Source	Source Vol (µl)	Cal* Diluent (µl)	Final Vol (µl)	Final Concentration (ng/ml)
1*	Cal. Int. (10µg/ml)	96	404	500	1920
2*	1*	50	50	100	960
3*	2*	50	50	100	480
4*	3*	50	50	100	240
5*	4*	50	50	100	120
6*	5*	50	50	100	60
7*	6*	50	50	100	30
Neg	-	-	100	100	0

1. Remove coated microtiter plate from -20°C and allow it to acclimate to room temperature for 5-10 minutes.
2. Dilute calibrators and test samples 1/100 with assay buffer (for example add 5µl of prepared calibrator or sample to 495µl of assay buffer). Mix well. Do not store diluted samples.

3. Add 100µL diluted calibrators and samples to appropriate wells on the plate. Incubate for 1 hour at room temperature on a plate shaker at 300rpm.
4. Discard the content of the plate and wash the wells 3x with 200µL wash buffer per well.
5. Add 100µL detection reagent to appropriate wells on the plate. Incubate for 1 hour at room temperature on a plate shaker at approx 300rpm.
6. Discard the content of the plate and wash the wells 3x with 200µL wash buffer per well.
7. 100µL of TMB to each well on plate. Incubate for 5 minutes at room temperature protected from light.
8. Add 100µL of TMB stop solution to each well on plate. Mix by gently tapping the side of the plate.
9. Determine absorbance with a microplate reader at 450nm against 620nm.

Calculations and results

1. Construct a standard curve by plotting the absorbance obtained from each standard against concentration. Use a 4 or 5 parameter curve fit.
2. The concentration of the unknowns can be back calculated directly from this standard curve using the absorbance value for each sample.

Performance characteristics

Precision: Precision was determined by analyzing samples at 800 (HQ), 300 (MQ), and 100 (LQ) with 6 replicates on 6 different occasions. Intra-assay and Inter-assay coefficient of variation (CV) was < 20%.

Cutpoint: Analysis of twenty treatment naive samples in the absence and presence of 50 ng/mL of somatotropin resulted in a floating cut point.

Detection Limit: The detection limit is 30 ng/mL.

Specificity: In presence of interfering materials, prolactin, IGF-I, IGFBP3, and rheumatoid factor (1 ng/mL) HQ and LQ met the acceptance criteria of having a percent error of ± 25%.